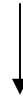


## How spectroscopic computations fit into the standard structural assignment protocols:

- ◆ To validate proposed assignments of spectral signals within a given structure (**signal assignment**)
- ◆ To evaluate the congruence of a structure to a set of experimental spectroscopic parameters (**structural determination**)
- ◆ To provide insight into the structural, electronic and environmental factors that influence the **spectroscopic parameters**
- ◆ To build **structure-property relationships**

# Fully theoretical prediction of the ESR spectra



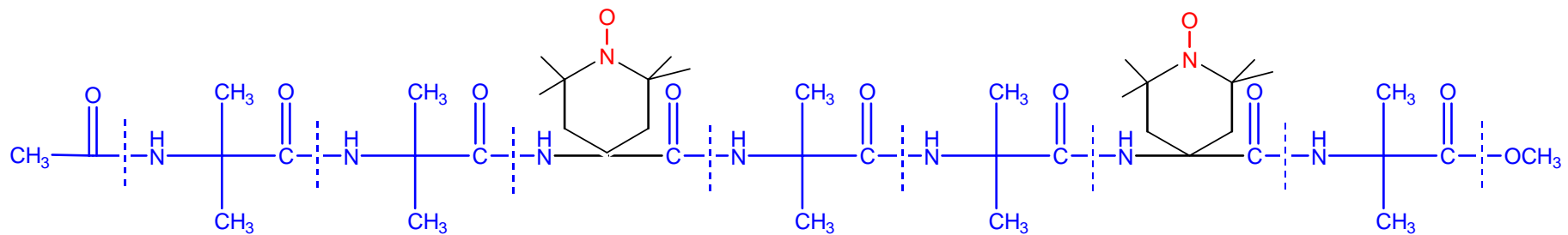
QM →  
geometric and magnetic observables ( $A_N$  and  $g$ -tensors) of the radical in its environment (DFT methods)

+

Direct feeding of calculated molecular parameters in a stochastic dynamic model (Stochastic Liouville Equation, SLE, formalism)

Integrated computational approach

Double spin labelled peptide:



ACE

AIB

AIB

TOAC

AIB

AIB

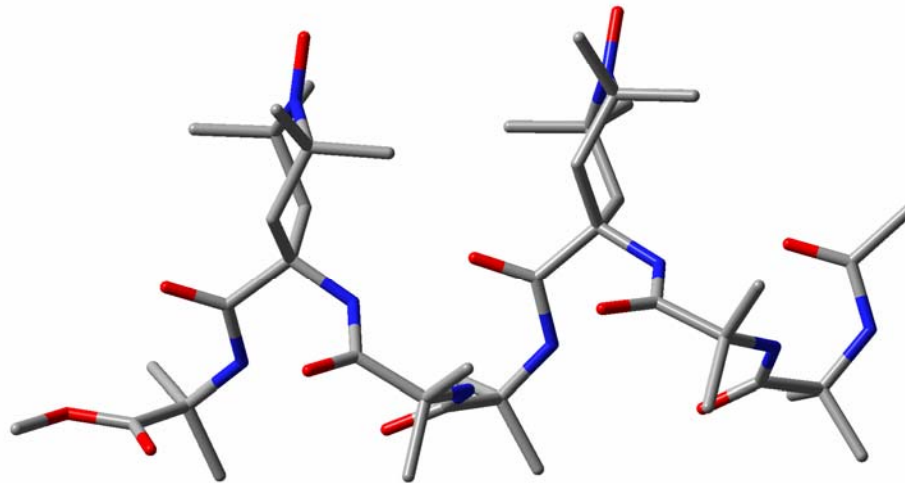
TOAC

AIB

OMe

## *Procedure:*

1. Optimization (DFT) of the structure [PBE0/6-31G(d) in CH<sub>3</sub>CN] → 3<sub>10</sub>-helical structure



2. Calculation DFT of the spectroscopic parameters (**A** and **g** tensors)
3. Evaluation of the diffusion tensor **D** and the dipolar interaction tensor **T**
4. ESR spectra simulation (Stochastic Liouville Equation)

## Stochastic Liouville Equation (SLE):

$$\frac{\partial \rho(Q, t)}{\partial t} = -i[\hat{H}(Q), \rho(Q, t)] - \hat{\Gamma}(Q)\rho(Q, t) = -\left[ i\hat{H}^\times(Q) + \hat{\Gamma}(Q) \right] \rho(Q, t) = -\hat{L}(Q)\rho(Q, t)$$

the time evolution of the density matrix of the system, depending upon general stochastic coordinates  $Q$ , controlled by the stochastic operator  $\Gamma$ .

$$\hat{H}^\times = \sum_{\mu} \sum_{l=0,2} \sum_{m, m'=-l}^l D_{mm'}^l(\Omega) F_{\mu, MF}^{(l, m')*} \hat{A}_{\mu, LF}^{(l, m)} \quad \hat{\Gamma} = D_x \hat{J}_x^2 + D_y \hat{J}_y^2 + D_z \hat{J}_z^2$$

The ESR spectrum is obtained as the Fourier-Laplace transform of the correlation function for the x-component of the magnetization:

$$|v\rangle = (2I + 1)^{-1} \left( |\hat{S}_{x,1}\rangle + |\hat{S}_{x,2}\rangle \right) \quad I = \text{nuclear spin}$$

$$I(\omega - \omega_0) = \frac{1}{\pi} \Re \langle v | [i(\omega - \omega_0) + (i\hat{H}^\times + \hat{\Gamma})]^{-1} | v P_{eq} \rangle$$

The diffusion tensor (**D**) was calculated for the rigid structure, considered as a set of spherical beads (i.e. extended atoms), was calculated via an “**hydrodynamic approach**”.

A friction tensor (**T**) is calculated via the comparison of the constrained and unconstrained molecular systems. The resulting tensor depends on the molecular geometry and solvent viscosity.

D is obtained as the inverse of T

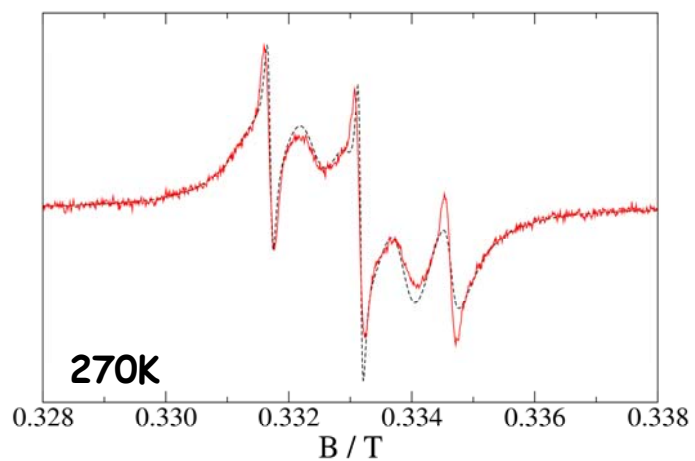
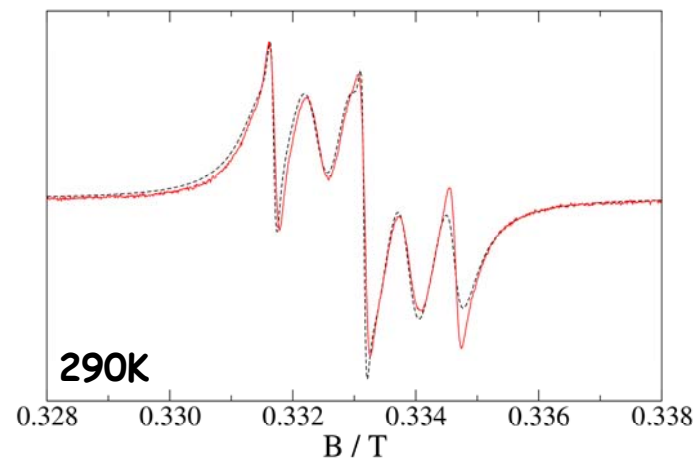
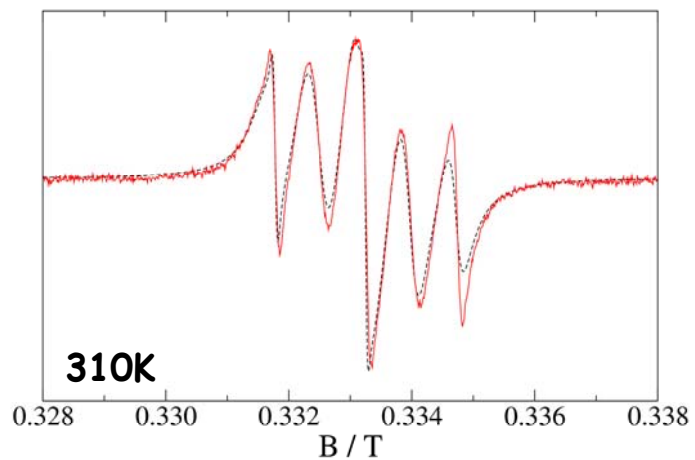
$$\mathbf{D} = k_B T \boldsymbol{\xi}^{-1} = \begin{pmatrix} \mathbf{D}_{TT} & \mathbf{D}_{TR} \\ \mathbf{D}_{TR}^{\text{tr}} & \mathbf{D}_{RR} \end{pmatrix}$$

---

Parameter	Source
$r_{12}$	6.6 Å/QM calculations
J	300 Gauss/Literature
D	Hydrodynamic model
T	Dipolar Interaction model
g	2.009, 2.006, 2.002/QM calculations
A	3.089, 3.325, 29.901 Gauss/QM calculations

---

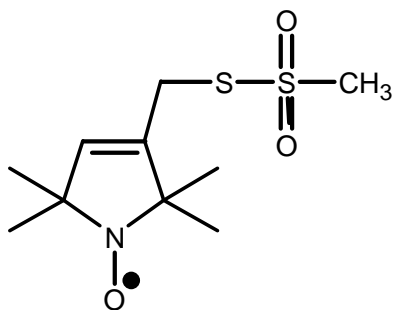
# ESR spectra of the eptapeptide in acetonitrile



Red lines: experimental spectra  
Black lines: simulated spectra

Computed values of g-tensor in gas-phase and in different protic solvents (water and methanol) and corresponding experimental data. Isotropic g value= $(g_{xx}+g_{yy}+g_{zz})/3$

Exp.l <sup>‡</sup>		TEMPO		TEMPO+1S		TEMPO+2S	
	MTSSL	<i>Gas-phase</i>	<i>Solution</i>	<i>Gas-phase</i>	<i>Solution</i>	<i>Gas-phase</i>	<i>Solution</i>
Methanol	<u>2.00574</u>	2.00624	2.00608	2.00589	2.00578	2.00563	2.00555
Water	<u>2.00551</u> $\Delta=0.00023$	2.00624	2.00607	2.00592	2.00580	2.00565	2.00559



(1-oxyl-2,2,5,5-tetramethyl pyrroline-3-methyl)  
methanesulfonate  
(MTSSL)

$$g = g_e + g_{RMC} + g_{DC} + g_{OZ/SOC}$$

$g_e$  = g value of the electron free (2.0023193)

$g_{RMC}$  = relativistic mass correction  $(-0.29196557 \cdot 10^{-3})$

$g_{DC}$  = diamagnetic correction to the g tensor

$g_{OZ/SOC}$  = Orbital Zeeman and Spin Orbit coupling contribution to g tensor

<sup>‡</sup>Owenius R. *et al.*, J.Phys.Chem.A, 2001, 105, 10967-10977.



*A combined qualitative analysis of experimental and calculated data make it possible to interpret the parameter shifts as due to the changed  $\epsilon$  and/or the increased propensity for hydrogen bonding.*

The integrated computational methods consisting of the most recent hybrid density functionals (DFT) and **mixed discrete-continuum solvent models** can be used to calculate shifts in  $A_N$  and g-tensor values due to changed dielectric (aprotic solvents) and hydrogen bonding properties (protic solvents), as well as the thermodynamic parameters.

CW-ESR Spectra of  
Fmoc-(Aib-Aib-TOAC)<sub>2</sub>-Aib-OMe

“Experimentally by **NMR** (in solution) and **X-Rays** measurements”



Secondary/Tertiary Structures of proteins and peptide?

Limit?

Investigation of :

- ◆ conformational transitions
- ◆ protein folding in real-time
- ◆ detailed study of dynamic properties of proteins in solution

Promising approach



SDLS  
Site-Directed Spin Labelling

+

ESR  
Electronic Spin Resonance

A combination of ESR spectroscopy

Double labeled  
systems...with two  
TOAC

ESR spectroscopy



- ◆ Structural parameters (distances..)
- ◆ Type of helical ( $3_{10}$  or  $\alpha$ )
- ◆ Coupling and dipolar interaction measurements

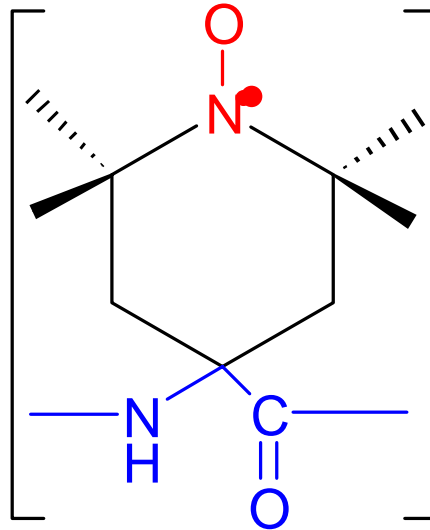
CW-ESR or ENDOR in SDSL



- ◆ Structural and dynamic informations

## TOAC

“artificial  $\alpha$ -aminoacid” which acts as a rigidly attached spin label in any chosen position of the peptide sequence

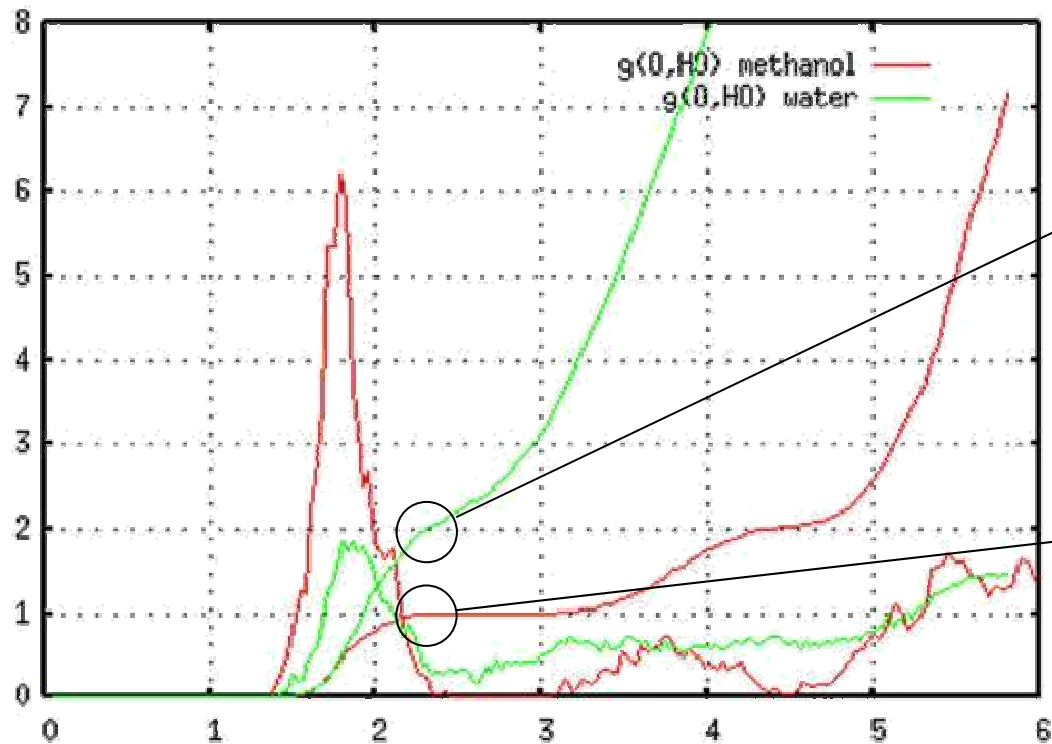


TOAC (4-ammino-2,2,6,6-tetramethylpiperidine-1-oxyl-4-carbossilic acid)

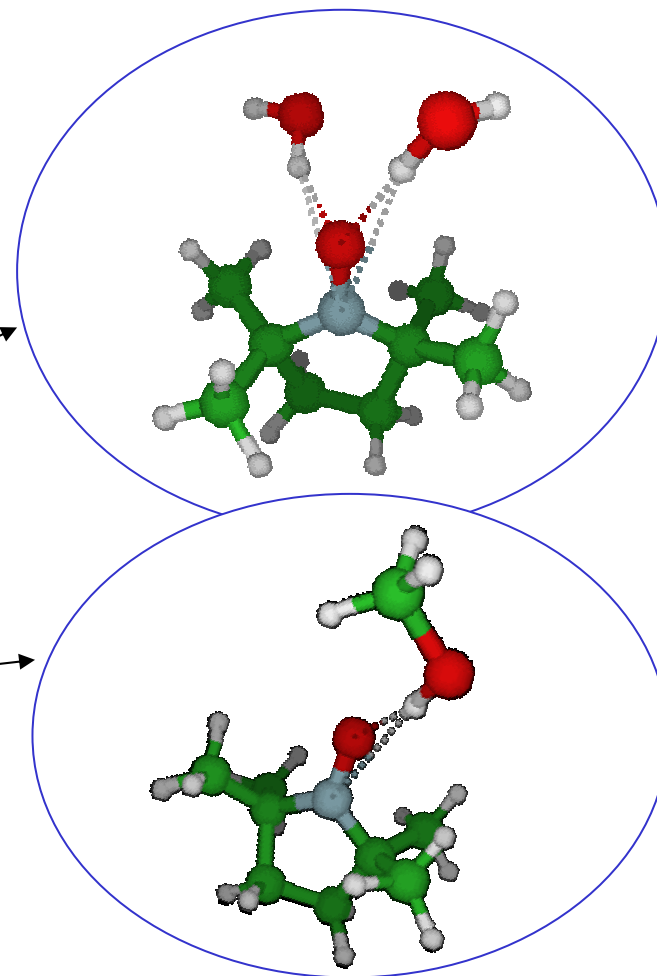
- ◆ TOAC belongs to the family of conformationally constrained  $C^{\alpha}$ -tetrasubstituted  $\alpha$ -aminoacids.
- ◆ TOAC is known to fold in a  $3_{10}$ -helical structure because the ring is rigidly attached to the backbone  $\alpha$ -carbon

# Car-Parrinello Molecular Dynamics (Proxyl)

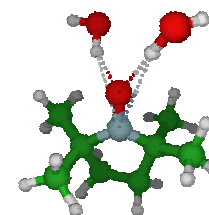
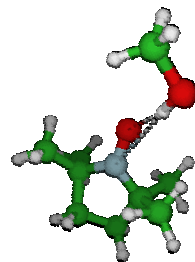
## Radial Distribution Functions (RDFs)



## SOLVATION SPHERE

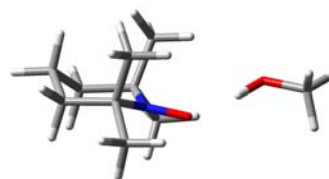


# Interaction Energies (kcal/mol)



	<u>Solvent-Solvent</u>		PROXYL+1S		PROXYL+2S	
	<i>Gas-phase</i>	<i>Solution</i>	<i>Gas-phase</i>	<i>Solution</i>	<i>Gas-phase</i>	<i>Solution</i>
Methanol	-5.9	-1.4	-7.7	-0.3	-14.4	<del>0.6</del>
Water	-5.7	-2.9	-7.9	-1.9	-15.6	-2.8

⇒ The interaction energies have been calculated in gas-phase on geometries optimized at PBE0/6-31+G(d,p) level. In solution they have been computed as single points at PCM/PBE0/6-31+G(d,p) level by UAHF model.



	<u>Solvent-Solvent</u>		TEMPO+1S		TEMPO+2S	
	<i>Gas-phase</i>	<i>Solution</i>	<i>Gas-phase</i>	<i>Solution</i>	<i>Gas-phase</i>	<i>Solution</i>
Methanol	-6.1	-1.7	-7.1	-0.7	-12.2	<del>0.3</del>
Water	-6.5	-3.5	-7.8	-2.5	-13.8	-3.8

⇒ The interaction energies have been calculated in gas-phase on geometries optimized at PBE0/6-31G(d) level. In solution they have been computed as single points at PCM/PBE0/EPR-II level by UAHF model.

## Hyperfine Coupling Constants ( $A_N$ in Gauss) of TEMPO



Exp.	TEMPO		TEMPO+1S		TEMPO+2S	
	<i>Gas-phase</i>	<i>Solution</i>	<i>Gas-phase</i>	<i>Solution</i>	<i>Gas-phase</i>	<i>Solution</i>
Methanol <span style="color: red;"><u>16.15</u> ‡</span>	14.94	15.75 (methanol $\epsilon=32$ )	15.51	16.14	16.06	-
Water <span style="color: blue;"><u>16.91</u> #</span>	14.94	15.80 (water $\epsilon=78$ )	15.50	16.10	16.03	16.50

⇒ The change in  $A_N$  can separately be interpreted in terms of the dielectric properties of the environment and the degree of hydrogen bonding.

⇒ The higher  $A_N$ -value in water might be explained by a larger number of hydrogen bonds formed in this solvent, since the influence of dielectric constant is very small in this region.

‡Aurich H.G.. *et al.*, Tetrahedron, 1977, 33, 969-975.

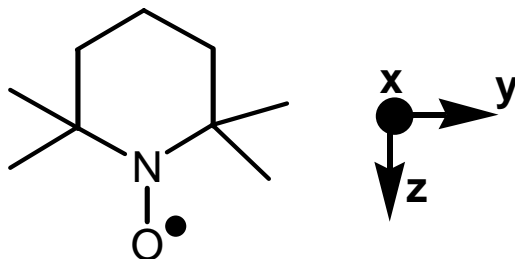
# Lim Y.Y. *et al.*, JACS, 1971, 93:4, 891-894.



## Calculated g-tensor of TEMPO in some solvents with different properties

Solvent	$g_{\text{iso}}$ $(g_{xx} + g_{yy} + g_{zz})/3$	$g_{xx}$	$g_{yy}$	$g_{zz}$	$\Delta g_{zz}$ (ppm)
Gas-phase	2.00624	2.00210	2.00638	2.01024	0.0
Cyclohexane	2.00621	2.00210	2.00637	2.01015	-91.7
Toluene	2.00620	2.00210	2.00636	2.01013	-110.2
Chloroform	2.00617	2.00210	2.00635	2.01007	-176.9
Isoquinoline	2.00615	2.00210	2.00634	2.01002	-224.0
Acetone	2.00614	2.00210	2.00634	2.01000	-246.9
Ethanol	2.00608	2.00210	2.00631	2.00983	-413.0
Methanol	2.00608	2.00210	2.00631	2.00984	-401.2
Water	2.00607	2.00210	2.00630	2.00982	-427.1

*Decrease*

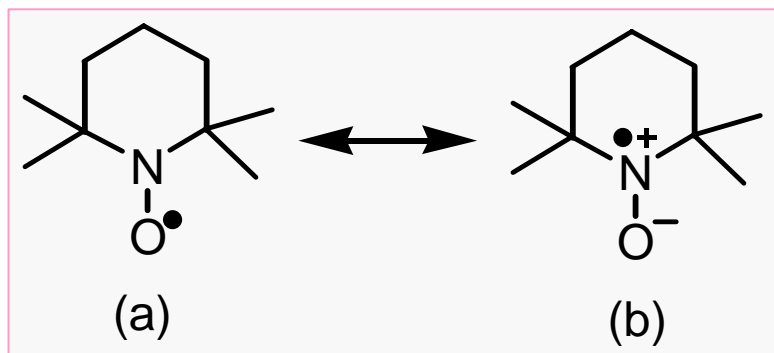


⇒  $g_{zz}$  is identified as the dominant contribution to the solvent dependence and decreases with increasing  $\epsilon$  of the solvent.

The g-tensor is calculated as a correction to the free electron value,  $g_e = 2.002319 \Rightarrow g = g_e + \Delta g$

$g_{xx}, g_{yy}, g_{zz}$ : diagonal elements of  $\mathbf{g}$ .

## Hyperfine Coupling Constants ( $A_N$ in Gauss)

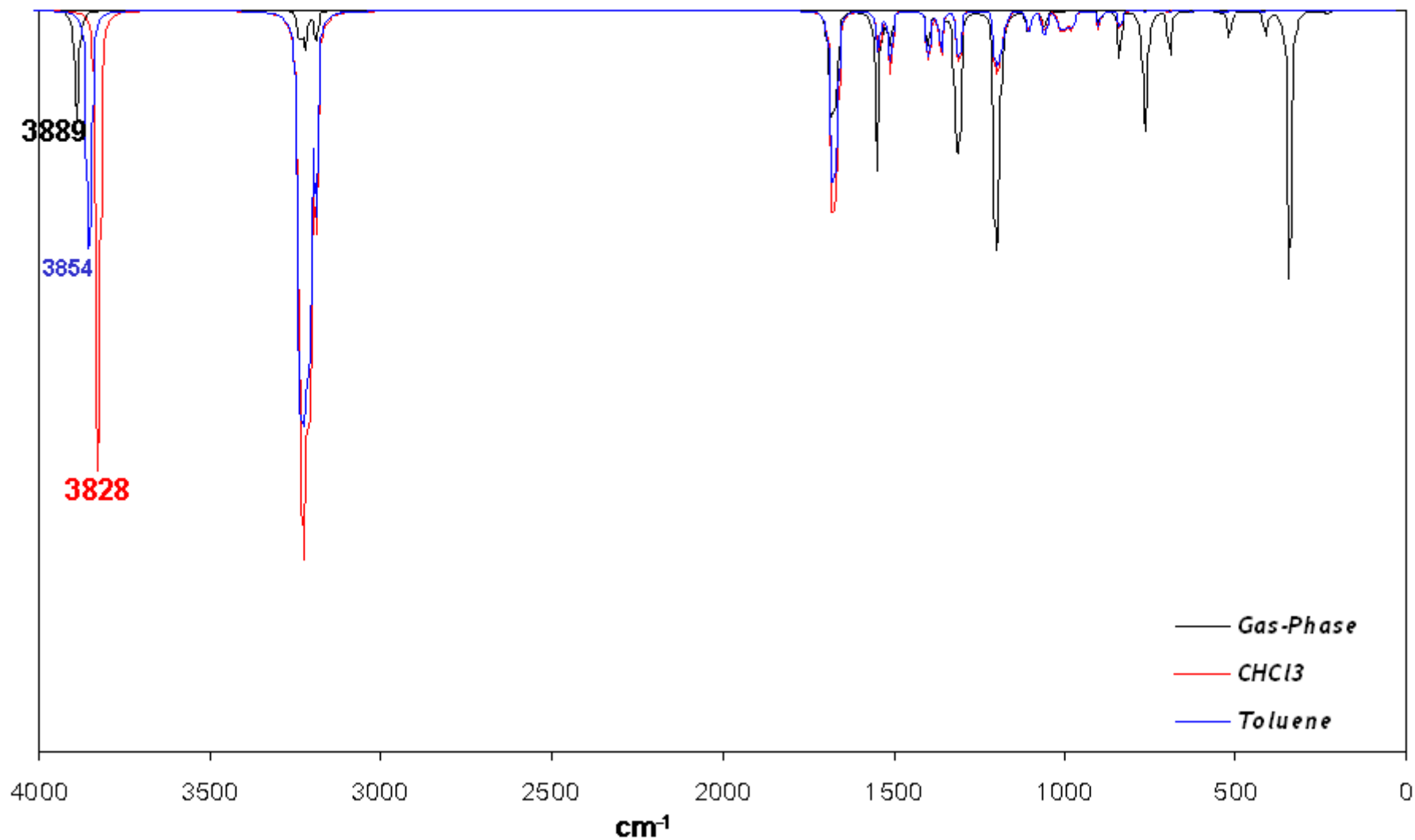


Polar solvent increase  $A_N$  through the relative stabilization of the “more polar” resonance structure

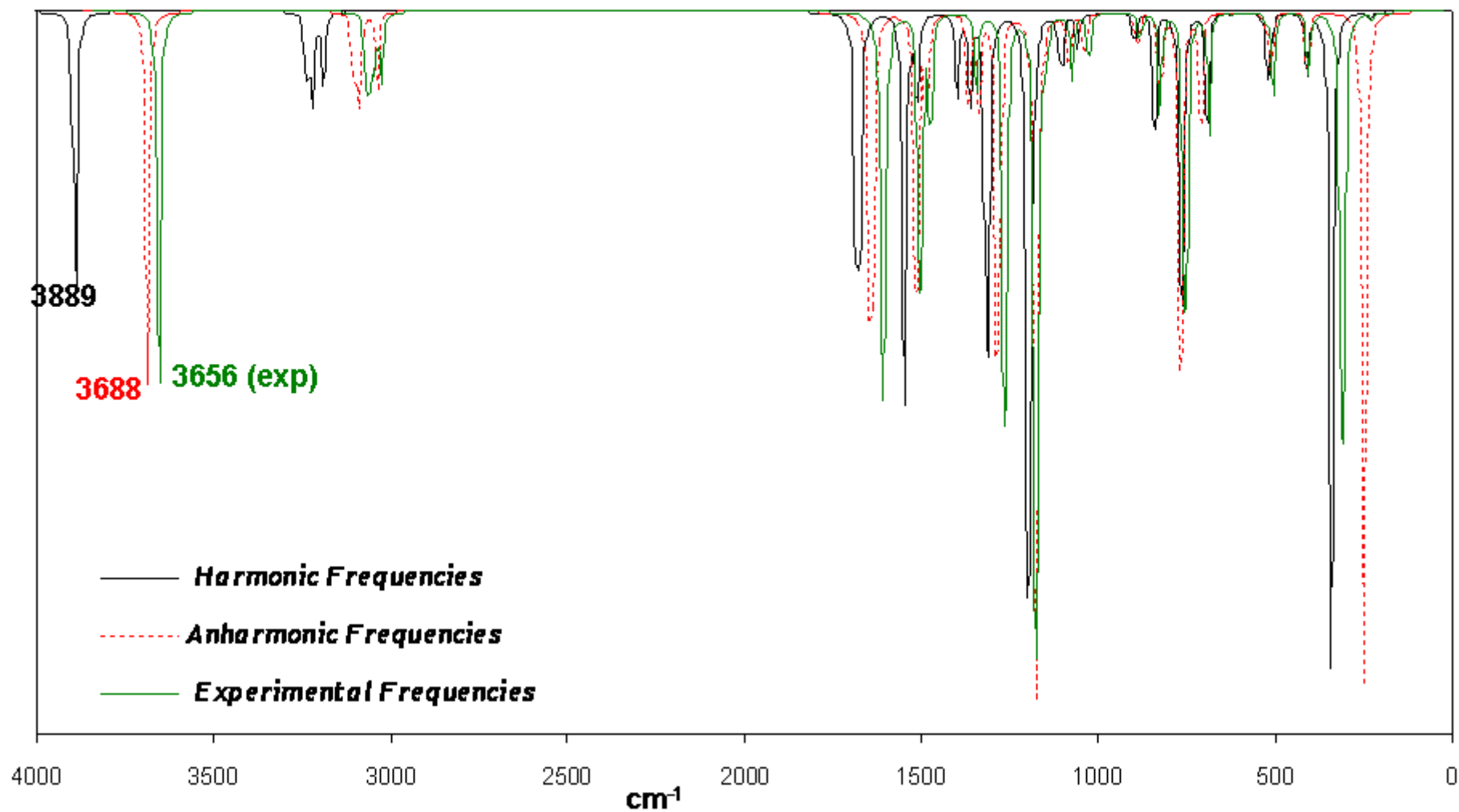
Exp	TEMPO		Bulk Effects +	Hydrogen Bond Effects	=	Overall Solvent Effects
	Gas-phase	Solution				
Benzylic Alcohol (cyclohexane)	<b>14.94</b>	<b>15.12</b> (cyclohexane $\epsilon=2.0$ )	<b>15.67</b> (benzylic alcohol $\epsilon=10$ )	<b>15.46</b>		<b>15.89</b> (benzylic alcohol $\epsilon=10$ )
Phenol (toluene)	<b>14.94</b>	<b>15.16</b> (toluene $\epsilon=2.4$ )	<b>15.70</b> (phenol $\epsilon=13$ )	<b>15.85</b>		<b>16.34</b> (phenol $\epsilon=13$ )

⇒ The  $A_N$  have been calculated in gas-phase as single points at **PBE0/EPR-II** on geometries optimized at **PBE0/6-31G (d)** level. In solution they have been computed as single points at **PCM/PBE0/EPR-II** level by UAHF model.

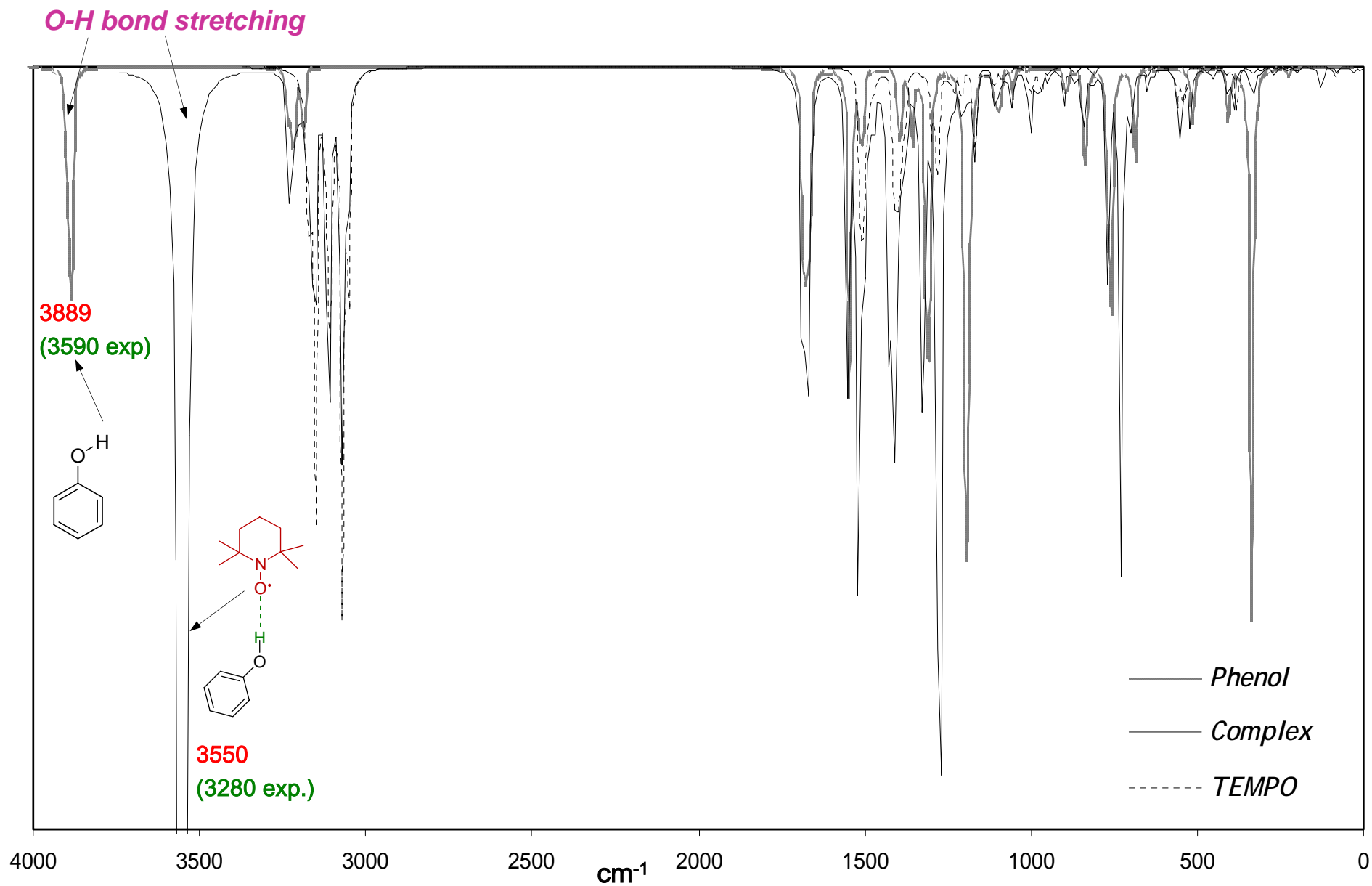
Computed FT-IR spectra for the free Phenol in gas-phase,  $\text{CHCl}_3$  and Toluene at PBE0/631+G(d,p) level and PCM/ PBE0/631+G(d,p) with UAHF model.



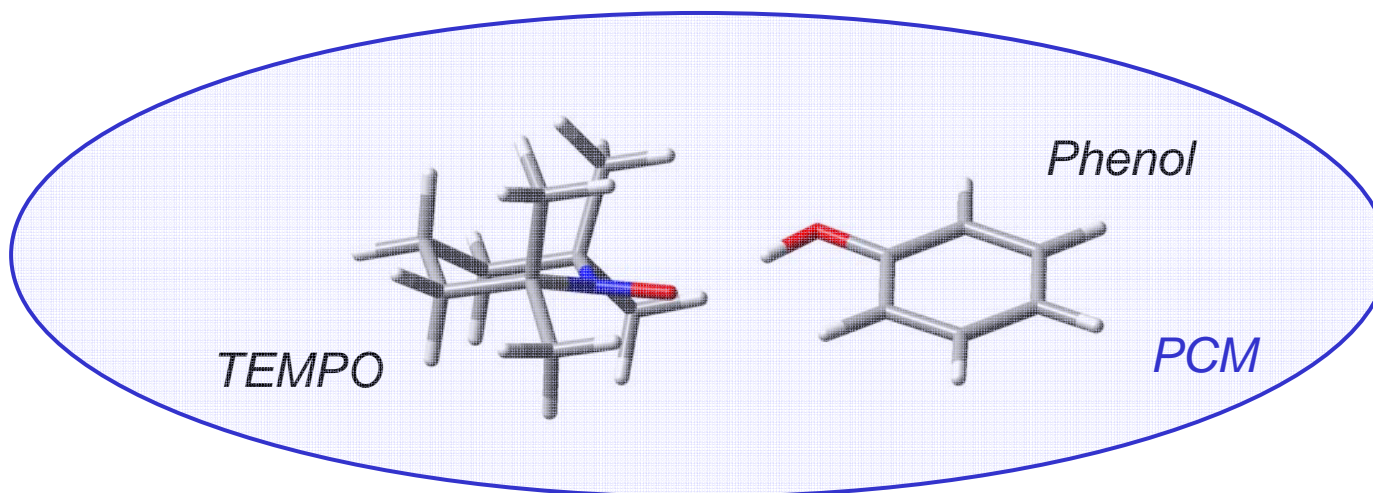
Experimental (gas-phase) and Computed FT-IR spectra for the  
free Phenol in gas-phase at PBE0/631+G(d,p) level.



Computed FT-IR spectra for the free phenol, TEMPO and hydrogen bond complex in gas-phase at PBE0/631+G(d,p) level.



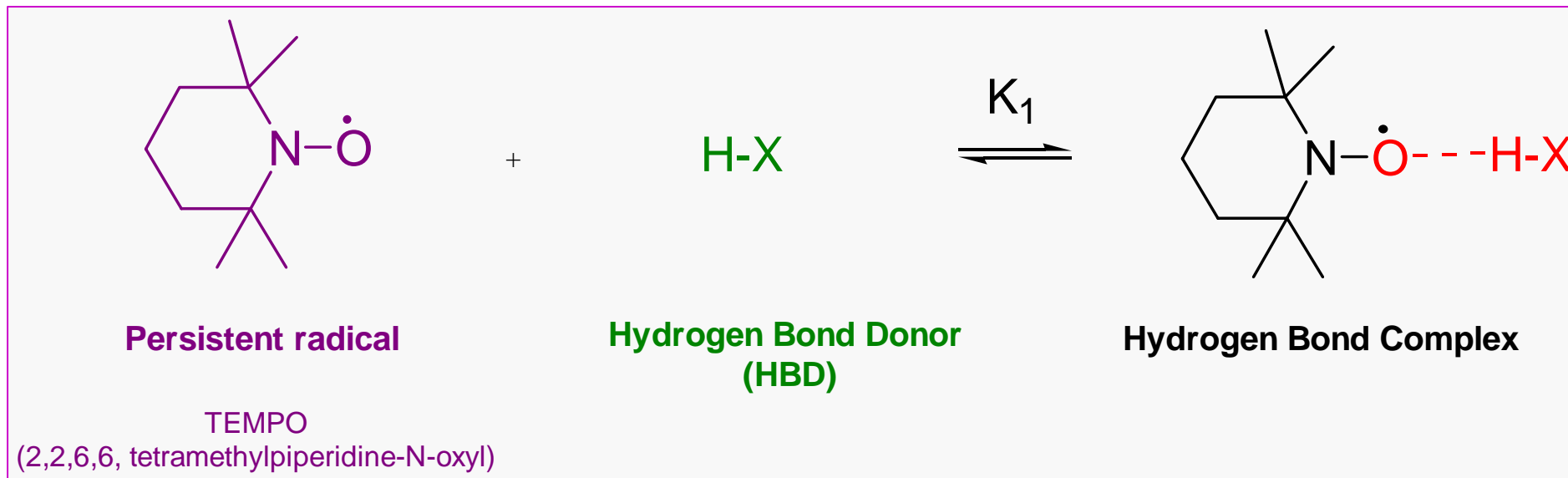
*Thermodynamic parameters* →  $\Delta H$



Computed vs Experimental

Gas-Phase	Solvent	Exp.
-6.03 kcal/mol	-4.91 kcal/mol	-4.69 kcal/mol

⇒ The formation energies have been calculated in gas-phase on geometries optimized at **PBE0/6-31+G (d,p)** level, and corrected for the basis set superposition error by the counterpoise method. In solution they have been computed as single points at **PCM/PBE0/6-31+G(d,p)** level by UAHF model.



→ Experimentally in solution by FT-IR and EPR measurements.<sup>‡</sup>

*Strength of the Hydrogen Bond*

→ Thermodynamic parameters ( $\Delta G$ ,  $\Delta H$ ,  $\Delta S$ )

→ Equilibrium constants ( $k_1$ )

→ Spectroscopic properties (Hyperfine Coupling Constant  $A_N$ ).

<sup>‡</sup>Pedulli *et al.*, CHEMPHYSCHEM, 2002, 3, 789-793.

*Study of Nitroxide Radicals in  
Condensed Phases*



Embedding medium (solvent) influences → *Energies, structures and properties*

***Solvent models***

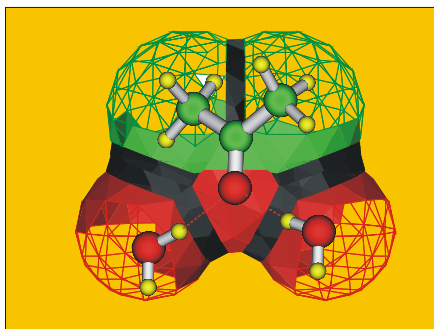
*Bulk Effects*

*Specific Effects*

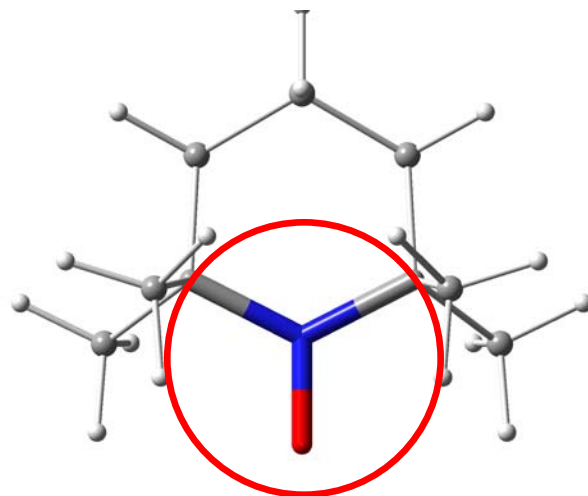
**Polarizable  
Continuum**

**Cluster**

*A mixed discrete/continuum model*



TEMPO



N-O = dimethyl nitroxide moiety  
(unpaired electron)



Once incorporated into the protein, spin label relaxation processes are dictated by molecular motion and local environmental properties.



ESR spectroscopic observables (A and g tensors)

Direct Information on solvent accessibility, topography of the polypeptide chain, electrostatic potential at any surface site, dynamic of the side chain, the distance from a second nitroxide and so on.

## The Spin Hamiltonian

$$H_s = \mu_B \mathbf{SgB} + \mathbf{SA} + \text{small terms} \quad \text{EPR}$$

- ✓ The **g**-tensor is calculated as a correction to the free electron value,  $g_e = 2.002319 \Rightarrow \mathbf{g} = g_e \mathbf{1} + \Delta \mathbf{g}$ .
- ✓  $\mu_B = \text{Bohr magneton} = eh/2m_e c$

**g**, **A** (hyperfine coupling tensor) can be expressed as **2<sup>nd</sup> derivatives** of the **energy** with respect to **the external field (B)** and/or **electron (S)/nuclear spin (I)**

$$\langle X \rangle = \frac{\partial^2 E}{\partial \lambda \partial \gamma}$$

For instance

$$\mathbf{g} = \frac{\partial^2 E}{\partial \mathbf{B} \partial \mathbf{S}}$$

➡ Find the terms of the electronic Hamiltonian that depends on external field and/or spin

# Hyperfine Coupling Constant ( $A=A_N1+A_{dip}$ )

Generally well reproduced by *ab-initio* methods for light atoms

- Fermi contact term

Non classical term: density at the nuclei

$$H_{iso} = -(2/3)g_e\gamma_e\gamma_N\mu_0\delta(\mathbf{r}_N)\mathbf{SI} \quad \text{Isotropic (A}_N\text{)}$$

Purposely taylored basis sets: EPR-II, EPR-III but 6-31+G(d,p) for structures and thermodynamic properties

- Dipolar term

Classical dipole-dipole interaction

$$H_{dip} = [(g_e\gamma_e\gamma_N\mu_0)/4\pi] [\mathbf{SI}/r_N^3 - 3(\mathbf{S}\mathbf{r}_N)(\mathbf{r}_N\mathbf{I})/r_N^5] \quad \text{Anisotropic (A}_{dip}\text{)}$$

In solvent with low viscosity (at ambient T that is true for all solvents in this study), the fast rotational motion of the spin label causes  $\mathbf{A}_{dip}$  to average out ( $\mathbf{A}_{dip}=0$ ).

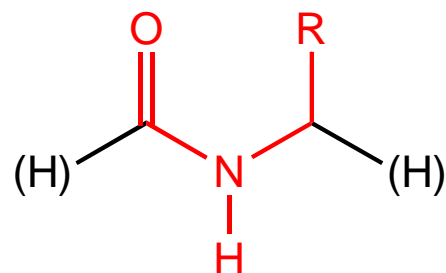
$\gamma_e$  and  $\gamma_N$ =magnetogyric ratios for electron and nucleus;  $\mu_0$ =vacuum permeability;  $\delta(r_N)$ =delta function which extracts the spin density at the nucleus;  $r_N$ =electron-nucleus distance.

Three different partition schemes (1-2-3) are tested within a QM/MM hybrid framework

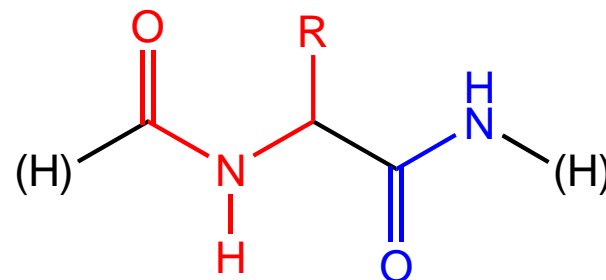
Inner → QM level

Atoms in the remaining part of the molecule → Point charges from AMBER parameters

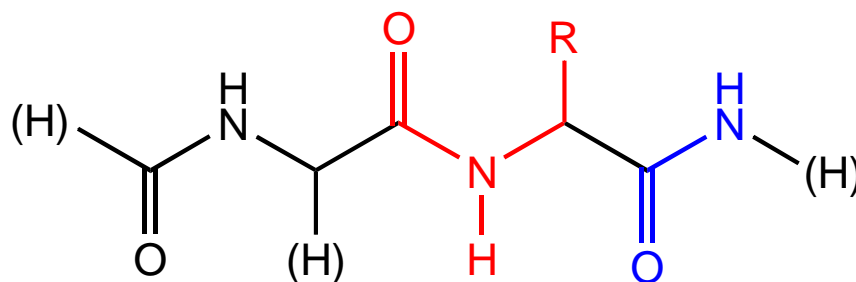
## QM regions: A1-A2-A3



**A1**



**A2**



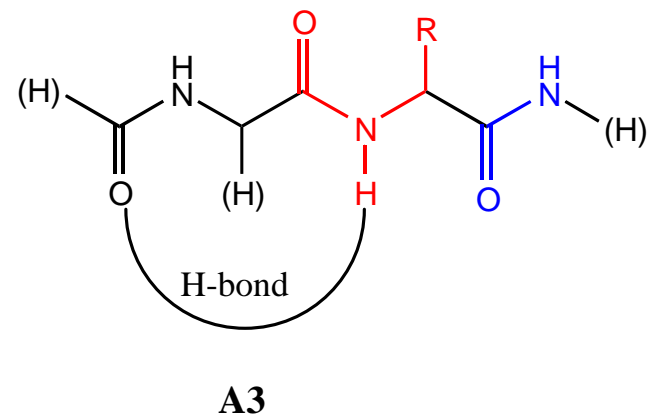
**A3**

Link atoms to saturate dangling bonds (to ensure that the model system has same electronic structure as the real system): H atoms for the broken bonds and the original bond distance is scaled in order to resemble a C-H bond length (0.7).

$\Delta_{2-3}$  = significant differences for  
**Ala2 , Gly4 and Gly8**

$\Delta_{1-2}$  = constant

Residue	1	2	3	BCP2 PBE0/ 6-311+G(2d,p)
Glu1	28.4	27.5	27.2	25.9
<b>Ala2</b>	26.6	<b>25.9</b>	<b>24.4</b>	22.8
<b>Gly4</b>	27.4	<b>27.4</b>	<b>26.6</b>	25.0
Lys5	29.2	28.2	27.8	26.0
Ala6	28.4	27.3	27.0	25.9
<b>Gly8</b>	27.0	<b>26.8</b>	<b>25.7</b>	24.0
Gly9	29.1	28.4	27.6	26.0



$$\sigma_{\text{iso}} = \sigma_{\text{iso}}(\text{MM}_{\text{all}}) + \sigma_{\text{iso}}(\text{DFT}_{\text{A3}}) - \sigma_{\text{iso}}(\text{MM}_{\text{A3}})$$

- ◆ QM region = fragment centered on the amide moiety of interest and including both peptidic bonds before and after the nucleus of interest.

The effect due to AMBER charges is significant only for those residues in which  $H^N$  is involved in hydrogen bonds (Ala2, Gly4 and Glu8), and for partition schemes 1 and 2, where the QM region is relatively small

- ◆ MM region = side chains have a minor influence on the chemical shift of amidic hydrogen atoms

Good agreement with full QM results !!!!

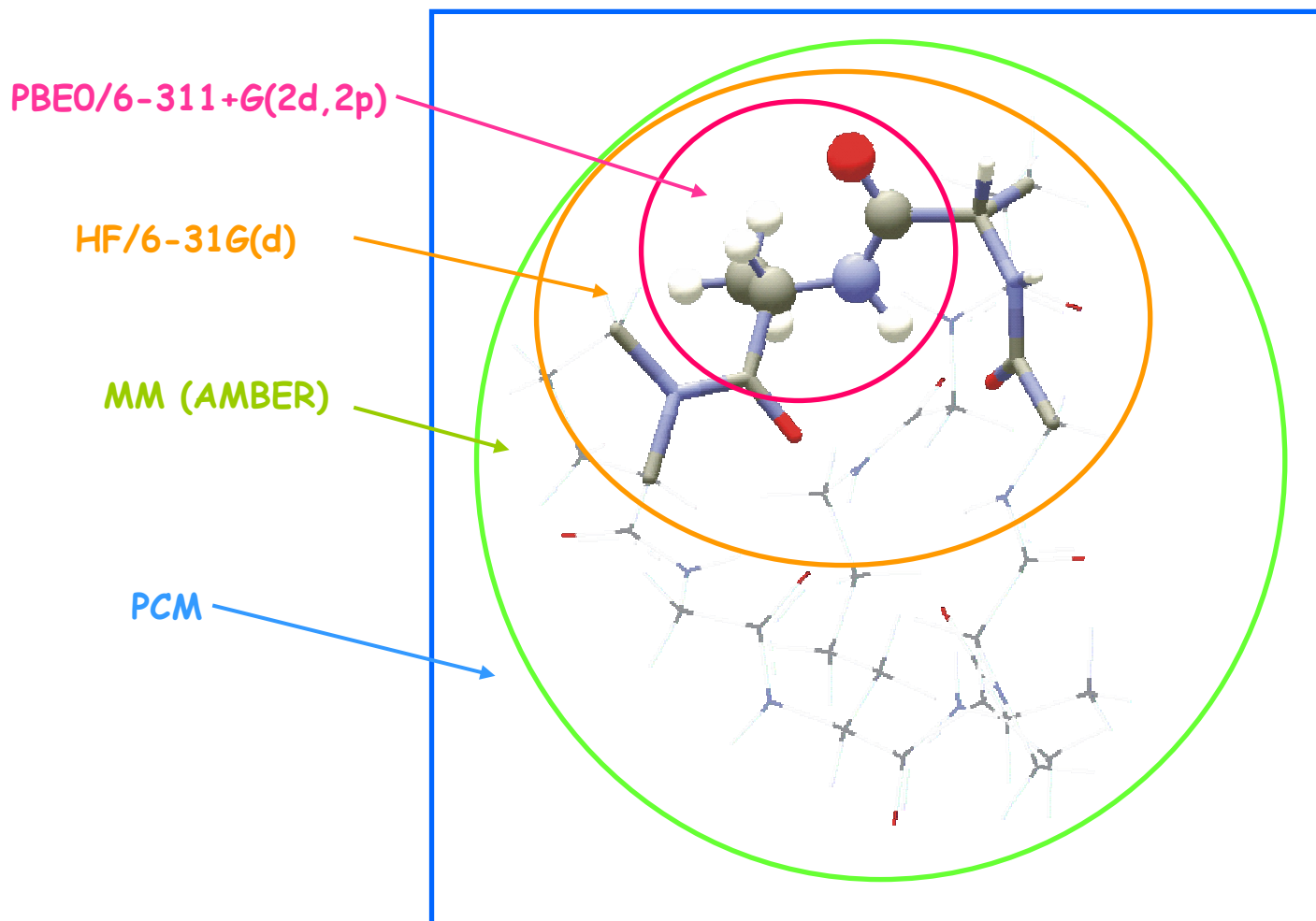


## Final three-layer QM/QM/MM hybrid method

Two different QM approaches are used in an ONIOM-like scheme

+

Point charges take into account polarization due to the remaining part of the molecule



This partition leads to a reduction in computer time of about one order of magnitude with respect to a two-layer (QM/MM) model and two orders with respect to a full QM model

## What do we want to compute?

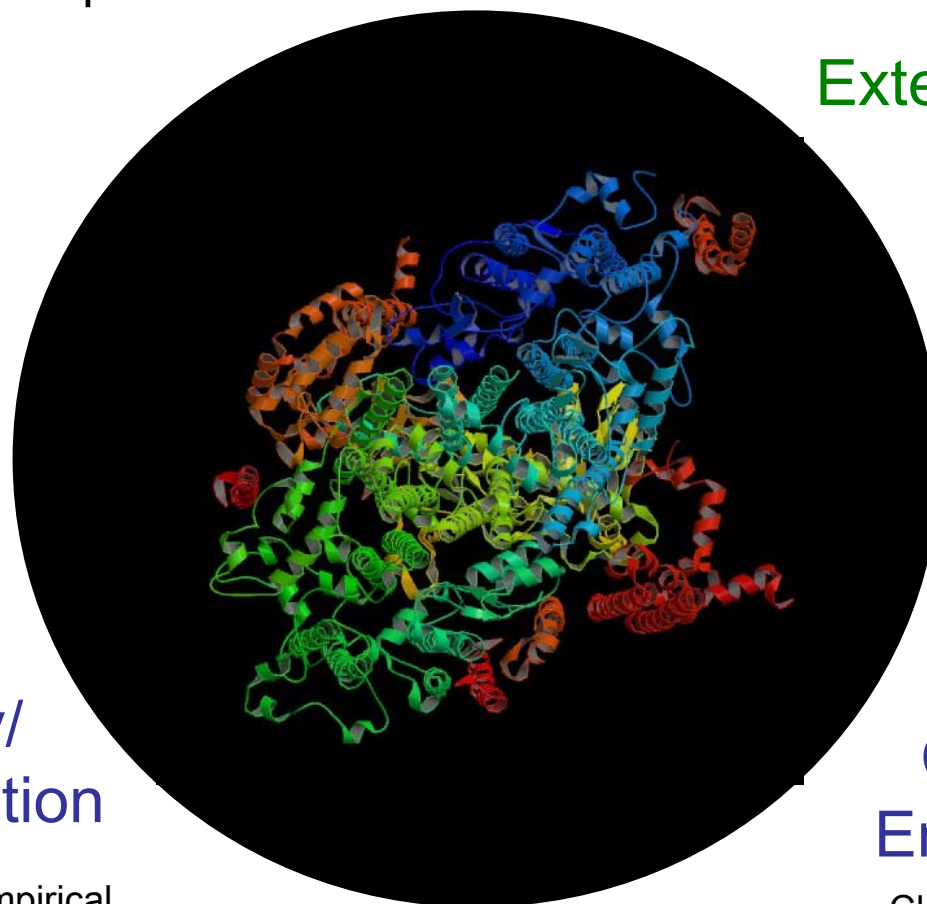
A property that depends on:

### Structure

Classical (MM, MD, ...)  
Quantum (structural minimization, CPMD, BOMD, ...)

### Spin density/ density distribution

Quantum: from semi-empirical to correlated methods



External field

Relativistic effects

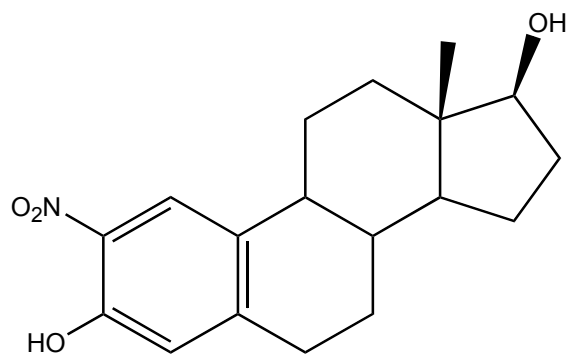
### Chemical Environment

Classical or quantum: embedding, dynamics, ...

## Main computational methods we use:

- **DFT** with hybrid functionals (**PBE0**): fast, amenable to treat large structures, or to explore significant portions of the conformational space (systematic conformational searches, grid searches etc.)
- For organic compounds, DFT works usually well for geometries and energies [**6-31G(d), 6-31+G(d,p)**], as well as for  $^1\text{H}$ ,  $^{13}\text{C}$  and  $^{15}\text{N}$  isotropic shifts [**GIAO, 6-311+G(d,p)**]
- **PCM** to model solvent medium; when specific interactions are involved (hydrogen-bonds), **cluster-PCM** approaches

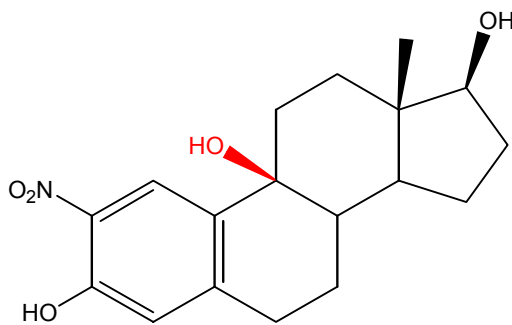
Calculation of  $^{13}\text{C}$  chemical shifts of  
nitroestradiol oxidation products



Precursor to compounds with anticancer activities and as probes for ligand-receptor interactions

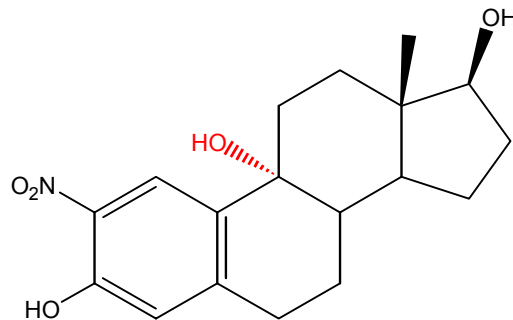
2-nitro-17β-estradiolo

Oxidation (peroxidase/H<sub>2</sub>O<sub>2</sub>)



**A**

(1:0.3)

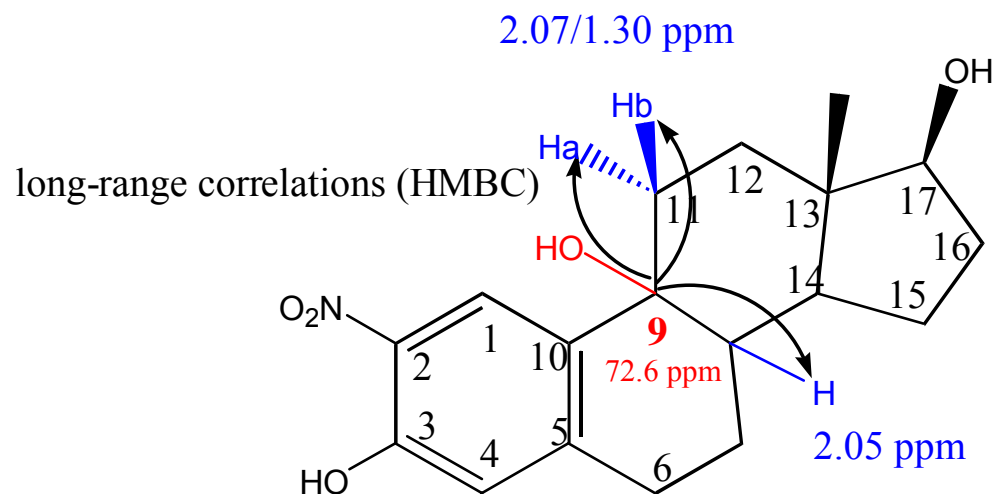


**B**

Very similar proton and carbon resonances

Isomers?

NMR experiments: <sup>1</sup>H, <sup>13</sup>C, <sup>1</sup>H COSY, <sup>13</sup>C HMQC, <sup>1</sup>H and <sup>13</sup>C HMBC  
UV-IR, MS-ESI



**A**

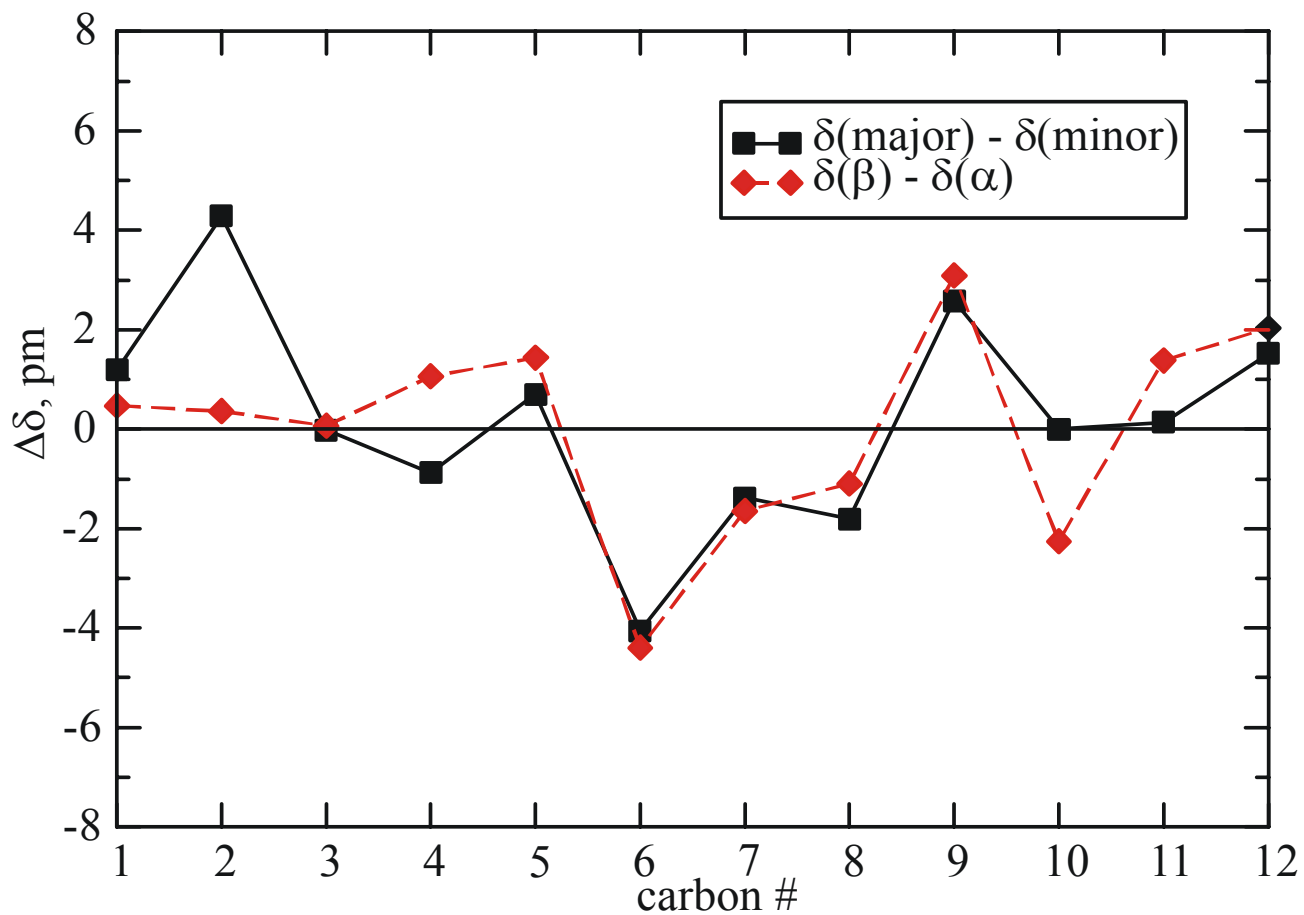
NMR analysis do not allow unambiguous stereochemical assignment to the distereoisomes



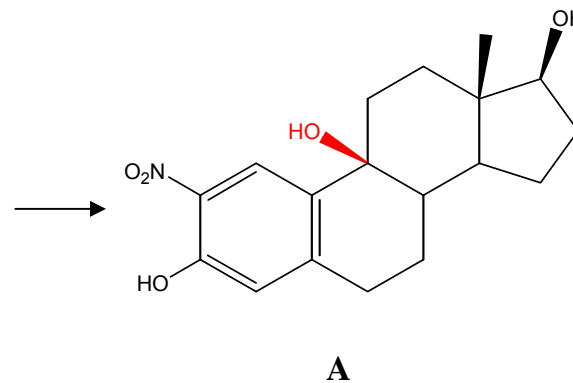
Quantum mechanical (QM) study on structures A and B (DFT )

Optimization PBE0/6-31G(d). NMR calculations PBE0/6-311+G(d,p).  
Solvent: acetone (PCM). Reference: TMS

Comparison of  $^{13}\text{C}$  chemical shift values calculated for the optimized structures of the  $9\alpha$  and  $9\beta$  configurations with experimental data



Assignment of the  $\beta$  configuration to the most abundant isomer (A)



**Calculation of amide proton  
chemical shift in a calcium binding  
protein**

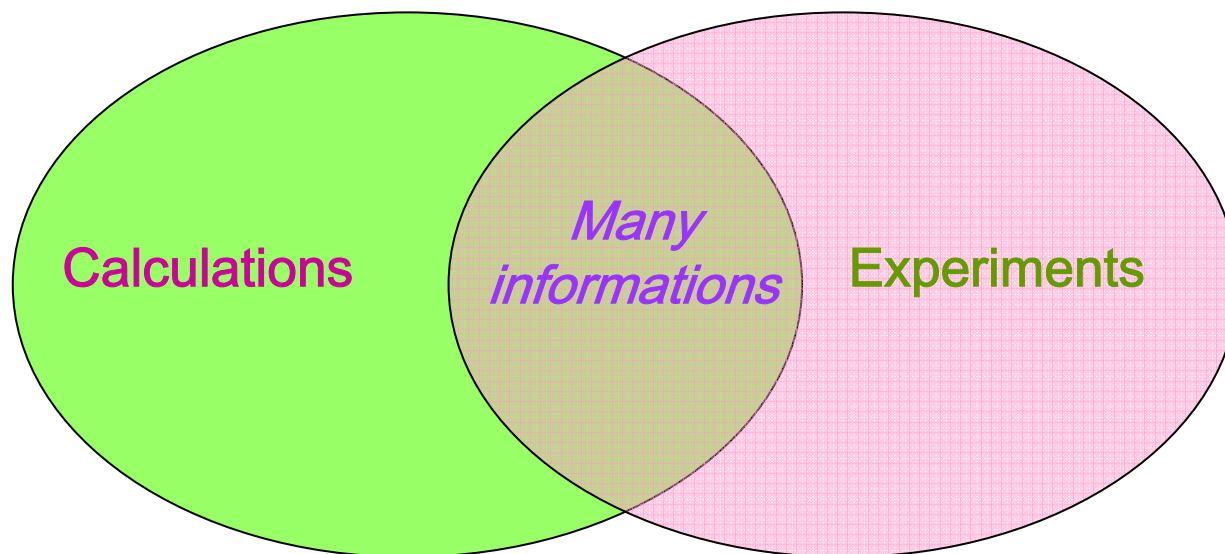


Chemical shift ( $\delta$ )  $\longleftrightarrow$  NMR (Nuclear Magnetic Resonance)

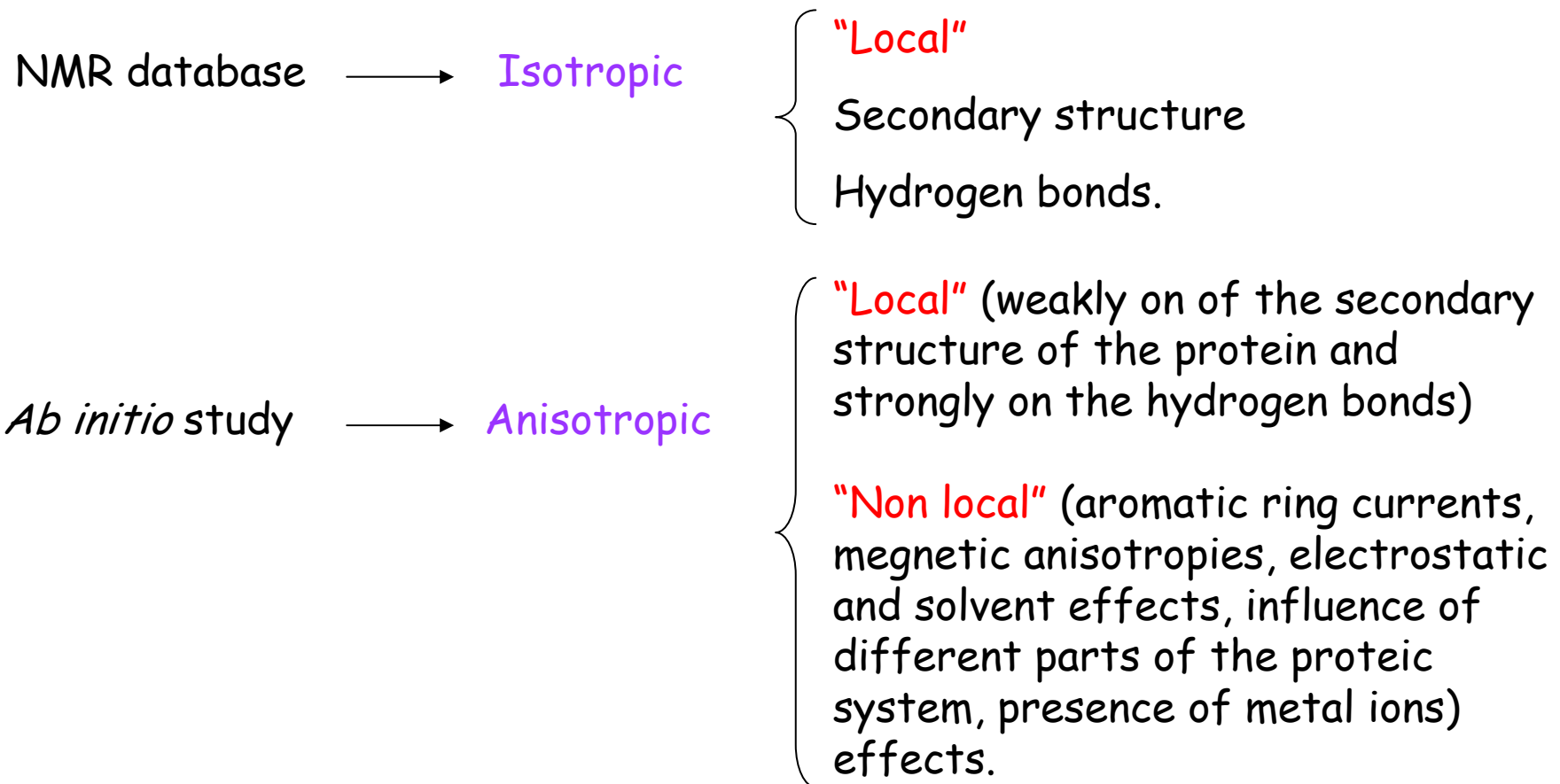
sensitive probe of molecular conformation, composition and environment.

widely used approach for studying the structure of chemical and biological systems.

Which is the relationship between chemical shifts and molecular structure ?



In peptides and proteins **amide protons ( $H^N$ )** are susceptible to additional local structural effects with respect to aliphatic protons.



## Which is the accurate computational model?

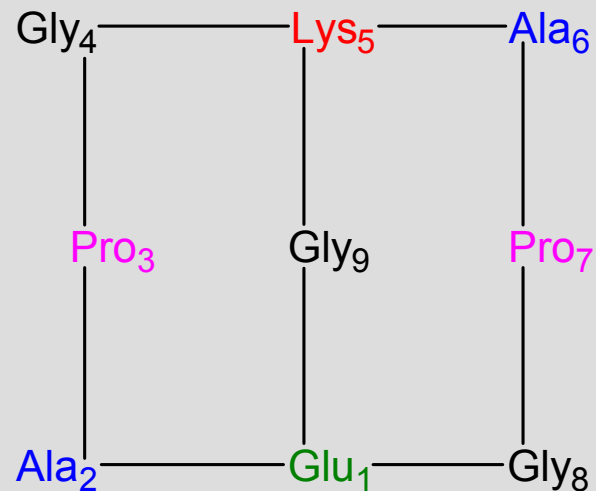
- ◆ A quite rigid peptide → local (structural) effects are of minor importance
- ◆ Right dimension of the system



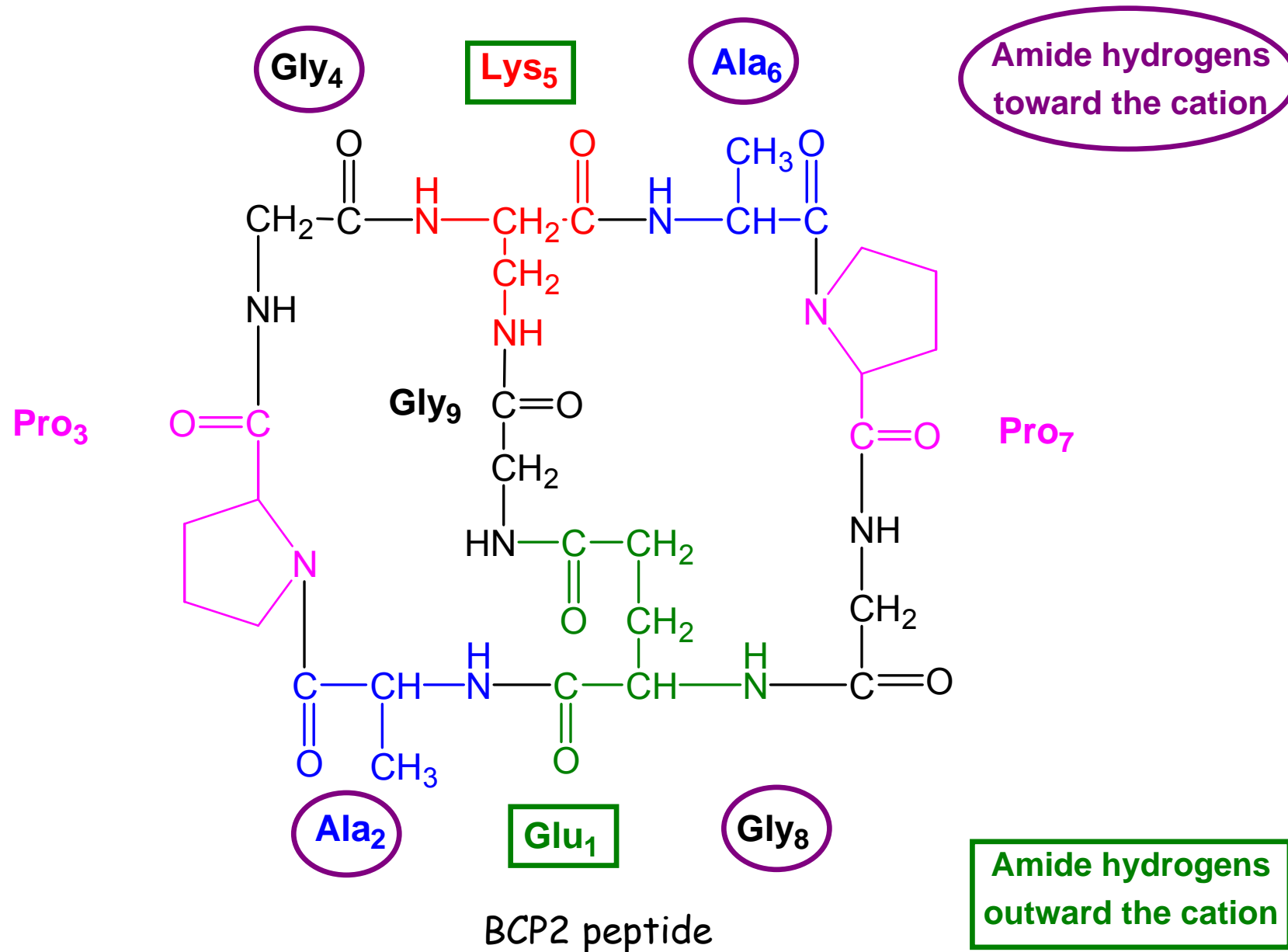
### BCP2

(bicyclic homodetic nanopeptide)

cyclo-(Glu<sup>1</sup>-Ala<sup>2</sup>-Pro<sup>3</sup>-Gly<sup>4</sup>-Lys<sup>5</sup>-Ala<sup>6</sup>-Pro<sup>7</sup>-Gly<sup>8</sup>)-cyclo-(1 $\gamma$ →5 $\epsilon$ )Gly<sup>9</sup>

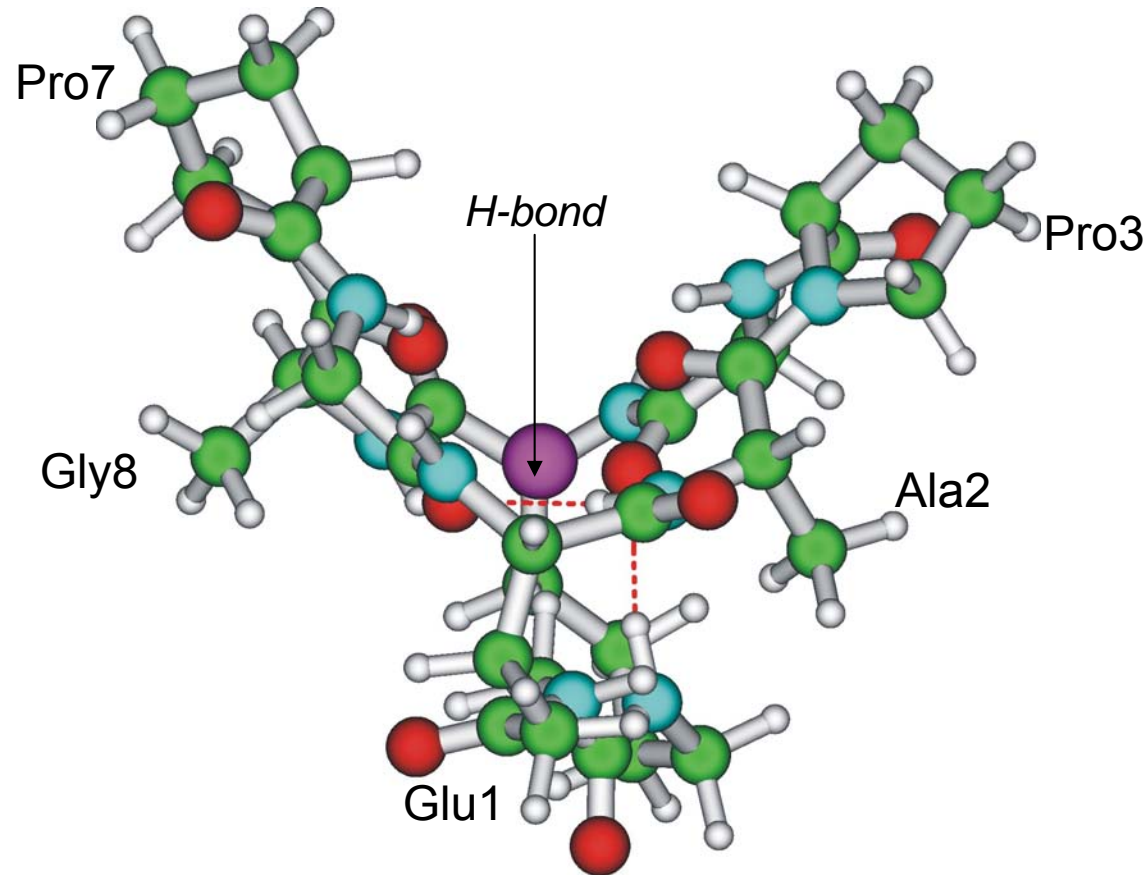


✓ A synthetic cyclic peptide: model for metal (calcium) binding polypeptides.



*...for the ab initio calculations*

1. Optimization (DFT) of the threedimensional structure experimentally determined



Model of the 1:1 calcium-BCP2 complex

The geometrical structure of the the calcium-BCP2<sup>+</sup> complex has been determined from NMR data followed by restrained molecular dynamics (RMD) calculations.

## 2. **Chemical shielding calculations** for the free peptide and the calcium BCP2<sup>+</sup> complex

$$\sigma_{ij}^N = [\partial^2 E / \partial B_i \partial m N_j]_{B=0}$$

HF and DFT level  
6-311+G(d,p) basis set

Chemical shift reference: acetamide

Chemical shift calculated + 6.40 ppm (i.e. the experimental proton shift of acetamide with respect TMS)

## 3. Calculations in solution (CH<sub>3</sub>CN)

Test with **PCM** → negligible solvent effect → **"isolated peptide"**

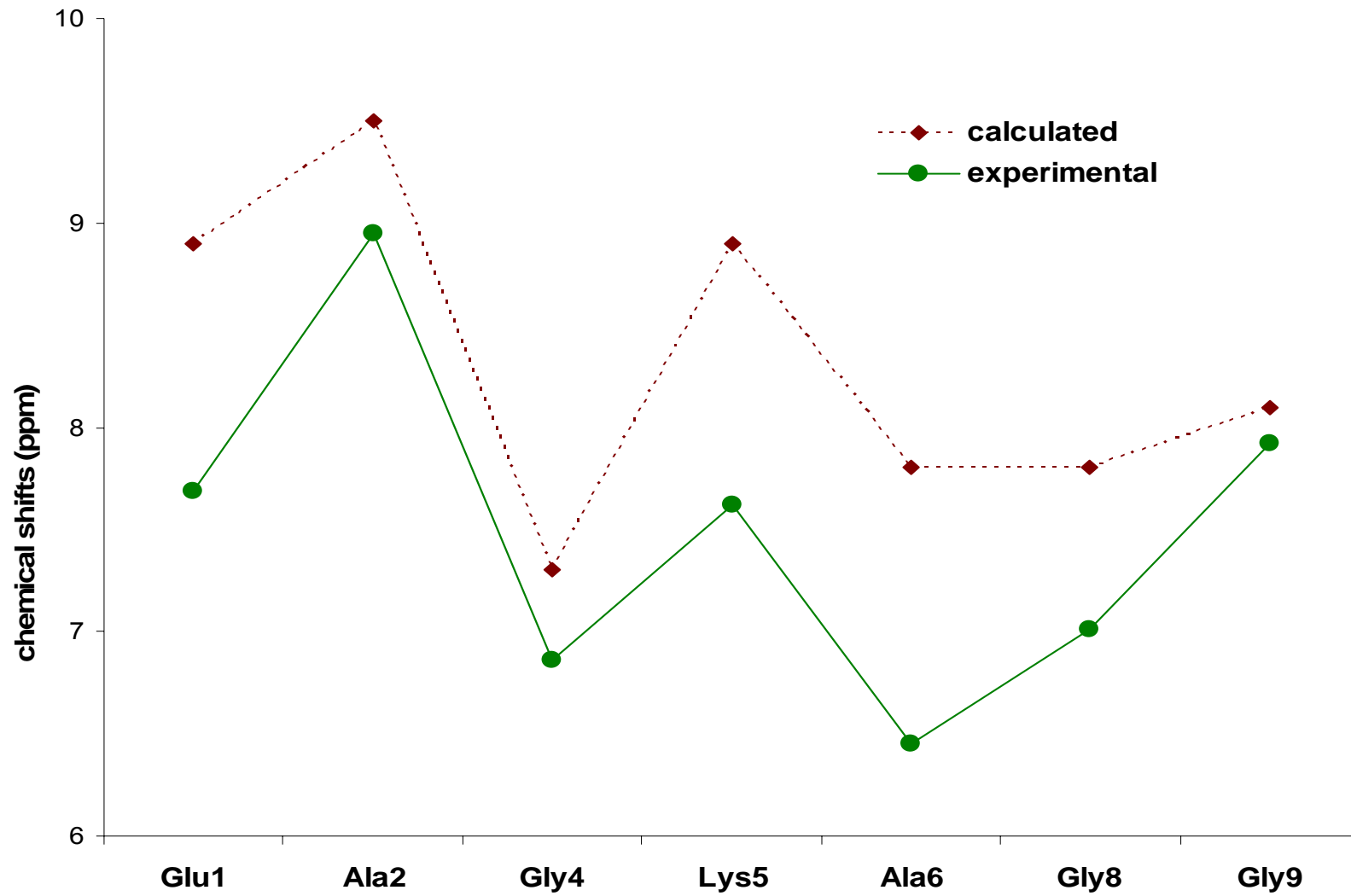
## Ca<sup>2+</sup>-BCP2 complex

	Glu1	Ala2	Gly4	Lys5	Ala6	Gly8	Gly9
DFT/6-311+G(2d,p)	8.9	9.5	7.3	8.9	7.8	7.8	8.1
HF/6-311+G(2d,p)	8.8	9.0	6.9	8.8	7.3	7.4	7.7
HF/6-31+G(d,p)	8.7	8.6	6.6	8.7	7.1	7.1	7.5
HF/6-31G(d)	8.8	8.5	6.6	8.9	7.1	7.0	7.7
HF/6-311+G(2d,p) for Ca <sup>2+</sup> and 6-31G(d) for all atoms	8.8	8.6	6.6	8.9	7.1	7.0	7.7
HF/6-31+G(2d,p) for N, O and 6-31G(d) for all atoms	8.6	8.8	6.6	8.6	7.1	7.2	7.5
<i>Experimental</i>	<i>7.69</i>	<i>8.95</i>	<i>7.62</i>	<i>7.62</i>	<i>6.45</i>	<i>7.01</i>	<i>7.92</i>

## Free BCP2

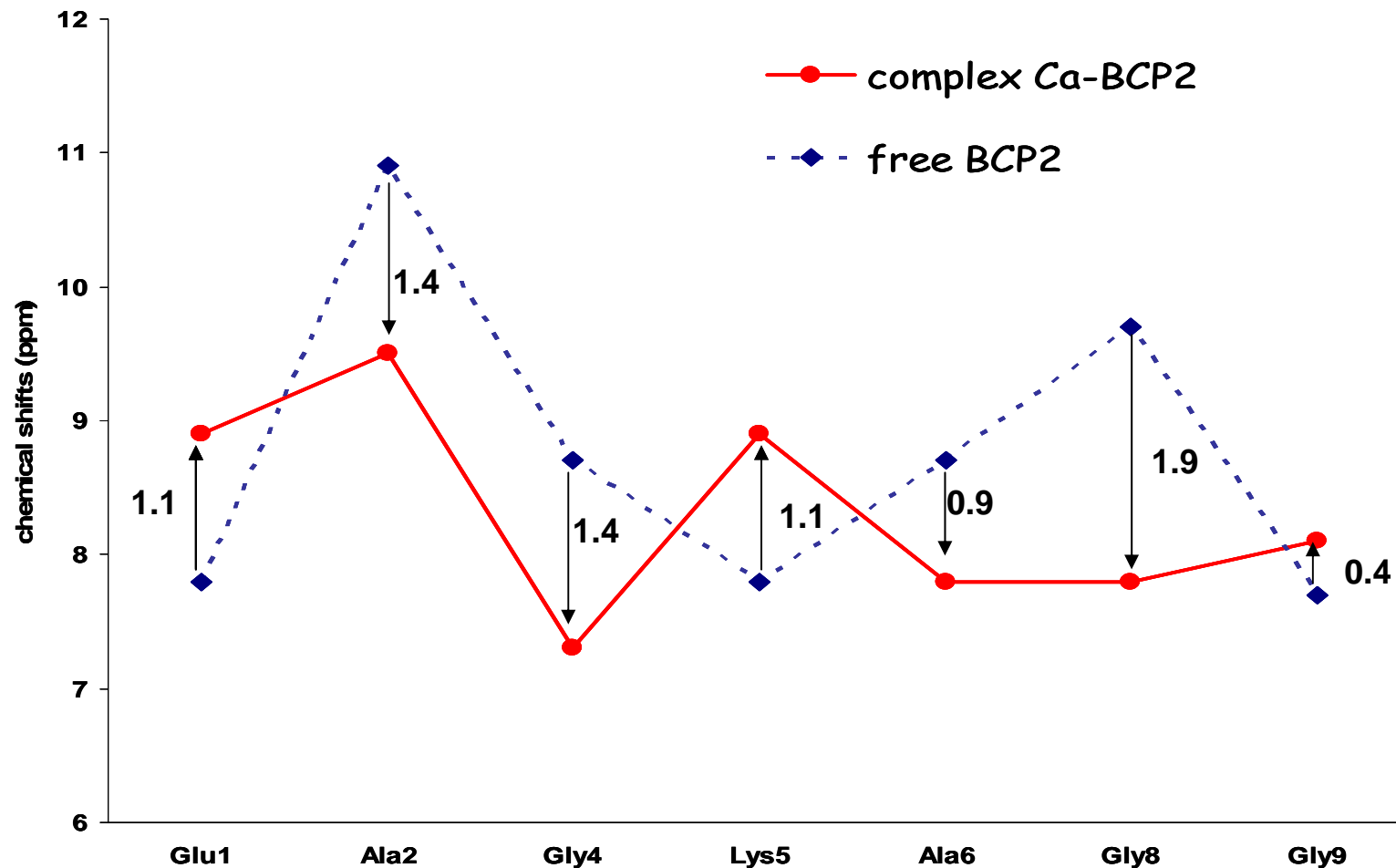
	Glu1	Ala2	Gly4	Lys5	Ala6	Gly8	Gly9
DFT/6-311+G(2d,p)	7.8	10.9	8.7	7.8	8.7	9.7	7.7
HF/6-311+G(2d,p)	7.4	10.7	8.5	7.4	8.3	9.6	7.3
HF/6-31G(d)	7.5	10.6	8.5	7.6	8.4	9.4	7.6

# Comparison of calculated [PBE0/6-311+G(2d,2p)] and experimental data for the Ca-BCP2 complex





# Comparison of calculated for the Ca-BCP2 complex and for the free peptide BCP2 [PBE0/6-311+G(2d,p)]



...focus on electronic effects due to the presence of the metal!!!!

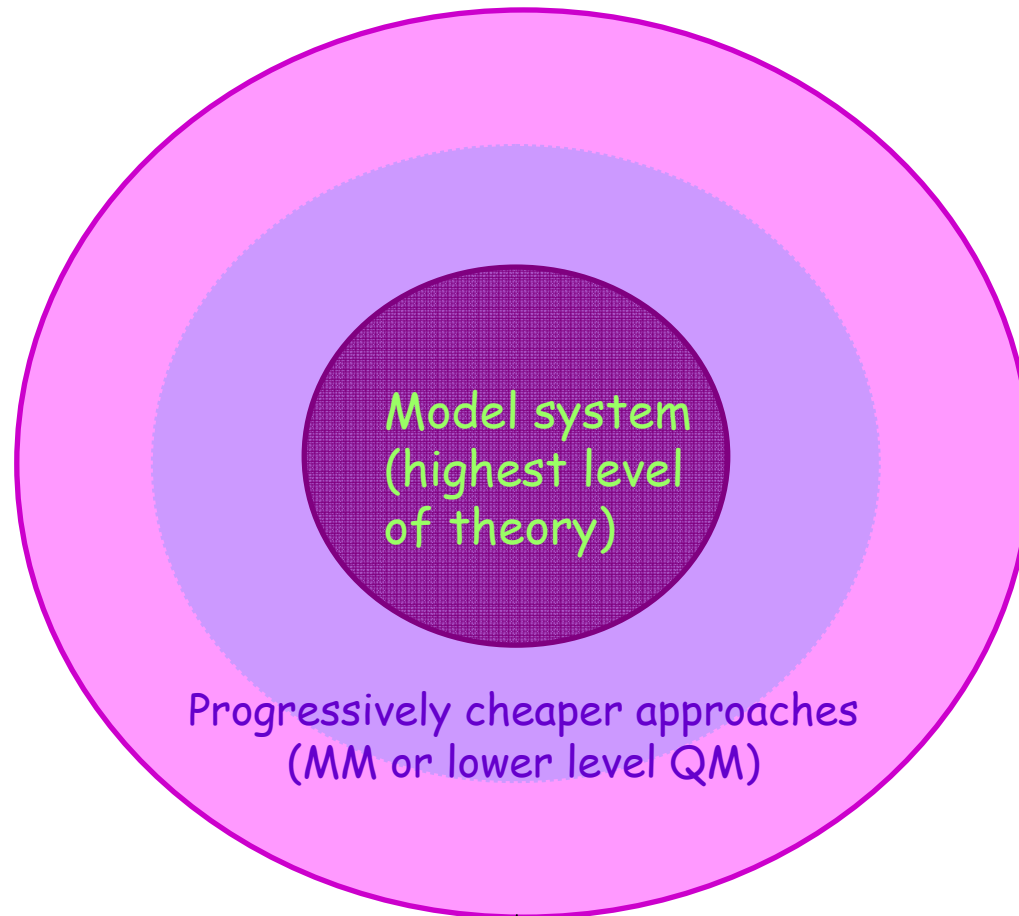
# First conclusions:

- ◆ Trends with different basis sets are in excellent agreement
- ◆ The use of an extended basis set limited to the metal cation is not sufficient for a good representation of metal-peptide interaction
- ◆ Relative shieldings values for amide protons (involved in the metal-peptide interaction) are almost equal in all calculations.

*“The effect of the cation is well reproduced”.*

# Hybrid Methods:

mixing QM and MM descriptions of different parts of the same molecule



Model system  
(highest level  
of theory)

Progressively cheaper approaches  
(MM or lower level QM)

MM region is described by atomic  
point charges.

1. Difficult parts of  
the molecule (those  
containing non  
parametrized atoms,  
uncommon interactions,  
or atoms directly  
participating in the  
transition state of  
reaction)

2. Parts of the  
molecule containing  
well parametrized  
atoms and simple  
interactions

3. Long range  
contributions on  
amide proton  
shieldings are  
mainly  
electrostatic

*...hybrid model*

What level (QM and MM) the ion and its ligands must be described?

✓ Test of different partition schemes  $\Rightarrow$  division of the system into QM and MM region trying to reproduce the full QM results

*1. Interaction between the cation and the peptidic chain*

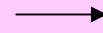
◆ The metal has to be treated at the quantum level

◆ Extended basis set reproduce better the charge transfer

*2. Long-range interactions*

◆ Electrostatic

*3. Special attention to hydrogen bonds*



Test: Calcium cation represented through different point charges, with charge +2.0 and +1.6 (chemical shifts calculated at HF/6-31G\* level)

↓

+2.0 in accordance with QM HF/6-31G\*  
+1.6 in accordance with QM/6-311+G(2d,p)